

Sample Size Requirements for Case-Control Study Designs.

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ABSTRACT

Published formulas for case-control designs provide sample sizes required to determine that a given disease-exposure odds ratio is significantly different from one. These are extended from one control per case to F controls per case and adjusted for a potential multi-category confounder in unmatched or matched designs. The effect of potential disease-exposure-confounder interaction may be explored. Interactive FORTRAN programs are described which compute the formulas.

Key Words: Biomedical computing; Confounder adjustment; Odds ratio; Power; Matched design.

1. Introduction

Breslow and Day [1] and Smith and Day [2] provide asymptotic formulas for the computation of case-control sample sizes required for odds ratios, unadjusted or adjusted for a confounder [1] and for matched designs [2]. The notation we use is their notation. Their formulas are extended here to include more than one control per case. The formulas for stratified matched sampling appear for the first time. They were deduced from applying the approach of Breslow and Day [1] (pages 305-6) to Table 7 of [2]. Modification of the formulas for specified interactions [1, 3] is also shown. These formulas are based on the logarithm of the odds ratio, for which the normal approximation is more accurate than for the exposure difference, so these formulas are more accurate than the exposure difference formula that is given in the majority of references [4, 5].

Two conversational FORTRAN programs, DAYSMITH and DESIGN, compute the formulas. They were submitted to STATLIB (lib.stat.cmu.edu) for non-commercial distribution a few years ago. The last known formula error was fixed in 1993. The programs produce a table of numbers of cases and controls required for a variety of specifications of Type I and Type II error, adjusted for the confounder, unadjusted, and adjusted for stratified matching, with the strata being the levels of the confounder. The two programs have different input requirements. Program DAYSMITH asks for exactly the items required for the Smith and Day formulas. Program DESIGN accepts alternative input that is converted in the program to the items required for the same formulas. The formulas used are shown in the Appendix.

2. The input to program DAYSMITH

The sample sizes computed are for the detection of a given disease-exposure odds ratio, that is, the sample sizes at which a certain statistical test will reject the null hypothesis that the odds ratio is one. The input items are as follows:

R_E = the odds ratio to be detected (typically a minimum value),

S = 1 or 2 for one-sided or two-sided type I error,

F = the number of controls per case,

P = the control population exposure probability, and

I = an indicator to request interaction adjustment.

Roughly speaking, interaction in statistics corresponds to effect modification in epidemiology. By not selecting an interaction adjustment, we effectively assume that the disease-exposure odds ratio does not differ across confounder levels. Interaction is discussed further in Section 6.

The number of confounder levels, denoted K is asked for next. If $K = 1$, unadjusted sample sizes only are computed, and no other input is required. Program DESIGN is identical to this point. For most applications, no confounder adjustment is required and so the program returns unadjusted sample sizes and is finished after a 1 is entered for K . The unadjusted formula [1] is more accurate than the usual unadjusted formulas [4, 5], and may therefore produce different sample sizes than those.

If $K > 1$, one of the levels of the confounder is taken to be a reference level, and is referred to as level one. The order of the levels is otherwise immaterial. The input required next is three numbers for each of the $K - 1$ remaining levels, p_{1i} , p_{2i} , and R_{Ci} , $i = 2, \dots, K$, which are

$p_{1i} = Pr(C_i|E)$ = among the exposed population, the proportion at level i of the confounder,

$p_{2i} = Pr(C_i|\bar{E})$ = among the unexposed population, the proportion at level i of the confounder, and

R_{Ci} = the disease-confounder odds ratio (with confounder level i versus level 1).

For the reference level, we set $R_{C1} = 1$ for the formulas that follow. We compute $p_{11} = 1 - \sum_{i=2}^k p_{1j}$ and $p_{21} = 1 - \sum_{i=2}^k p_{2j}$.

3. Input for Program DESIGN

Whereas DAYSMITH asks for the same input as requested in the original references [1-3], we found that alternative input made more sense for our initial applications [6, 7], so a second program was written. The input for DESIGN is the same as for DAYSMITH up to the point after which the number of levels of the confounder, K , is asked for.

Again, one of the levels of the confounder is taken to be a reference level, and is referred to as level one. The input that is required next is one number for the reference level, r_1 , and then three (four when interaction is included) numbers for each of the $K - 1$ remaining levels, r_i , p_i , and R_{Ci} , $i = 2, \dots, K$, which are

$r_i = Pr(E|C_i)$ = the probability of exposure at level i of the confounder,

$p_i = Pr(C_i)$ = the probability of being in level i of the confounder, and
 R_{C_i} = the odds ratio of disease and confounder level i (versus level 1).

For the reference level, we again set $R_{C_1} = 1$.

From Bayes Theorem, we compute

$$p_{1i} = r_i p_i / P \quad \text{and}$$

$$p_{2i} = (1 - r_i) p_i / (1 - P).$$

We have one more input item than is actually required, and that is used for a check, where we can use the fact that

$$P = \sum_{i=1}^K r_i p_i .$$

What we actually do is check the sum

$$\sum_{i=1}^K p_{1i} = \Delta .$$

The sum Δ is supposed to be equal to one. If it is not one, then we re-define and report $p_{11} = 1 - \sum_{i=2}^K p_{1i}$ and $p_{21} = 1 - \sum_{i=2}^K p_{2i}$, unless they are negative. An alternative used in earlier versions was to compute

$$\tilde{p}_{1i} = r_i p_i / P \Delta \quad \text{and}$$

$$\tilde{p}_{2i} = (1 - r_i / \Delta) p_i / (1 - P)$$

and replace p_{ji} with \tilde{p}_{ji} , for $j = 1, 2$ and $i = 1, \dots, K$. This is equivalent to replacing r_i with $\tilde{r}_i = r_i / \Delta$, $i = 1, \dots, K$, which is how the program used to report the change.

4. An example, adjusting for a confounder

The following example is one of several computations performed for a published research protocol for a study of the association of oral contraceptive (OC) use with cardiovascular risks, controlling for age group [6]. A related protocol [7] has smoking as a confounder.

The numbers entered for P , r_i , p_i , and R_{C_i} , $i = 2, \dots, K$, are all taken from the Saskatchewan government medical database, which includes the entire population from which a case-control sample is to be taken. In many applications, such numbers are not available from a reliable source. In that case, one may try sets of alternative minimum and maximum

numbers for a range of results. The maximum sample sizes obtained from such sensitivity analyses would be the conservative recommendation.

Both programs first request R_E to I . For R_E , the outcome of interest is hospitalisation due to certain cardiovascular risks. The exposure is a specific OC with 10% of the market share [7]. Since overall OC prevalence is 30%, then $P = .03$ for that specific OC. Using $>$ to denote the cursor for computer entry, we type:

```
>2 2 3 .03 0
```

for R_E , S , F , P and I , respectively, then press enter. We then receive the message:

Type the number of confounder levels, and <enter>. Type 1 if no confounder.

We enter 5 levels and press enter.

```
>5
```

Now type in the population exposure probability for the reference level of the confounding variable.

This will be put at level 1, so it is $\Pr(E|C_1)$

The confounder levels are five age groups, and level 1 corresponds to the youngest age group 15-21, for which we enter the prevalence for a specific OC with 10% of the market share. We type .055 and press enter.

```
> .055
```

The reply is:

Now type in, for each of the other 4 level(s) of the confounding variable, $\Pr(E|C_i)$, $\Pr(C_i)$, and $R_c(i)$, separated by at least one blank or <enter>, where $\Pr(E|C_i)$ = in the population at level i of the confounder, the proportion exposed, $\Pr(C_i)$ = the probability of being at level i ,

and $R_c(i)$ = odds ratio of disease and confounder level i (versus level 1).

The following numbers are entered for age groups 22-26, 27-31, 32-39 and 40+:

```
> .038 .24 2
```

```
> .021 .2 8
```

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> .008 .18 8
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```
> .004 .15 28.5
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Note that $Rc(5) = R_{C5} = 28.5$, a very high value. That is to be expected if all older women are included. (For the final protocol [6], a cut-off was made at age 45.) When enter is pressed, we receive some confirmation of the input, and a message that the result is written to file design.out. That is, as currently written, the sample sizes and other output are not automatically shown on the screen, but are saved in “design.out” to be viewed directly there. Appendix 2 shows the output from the preceding session, which includes a correction of the input values.

Looking at Appendix 2, we see unadjusted sample sizes, those adjusted for age in an unmatched study, and a third set of sample sizes for a matched case-control study. For our example [6], both unmatched and matched designs are considered. With the low value of P and the high value R_{C5} , we see that a large difference in sample sizes required for either design may result. In most applications, however, the differences are not so dramatic.

5. Adjusting for a matching confounder

Epidemiological literature has to date usually given formulas for matching which are based on the strong assumption that all sources of extraneous variation among a case and its controls are accounted for [1, 8, 9]. A third program DESIGNM was written to compute such a formula (from [1], p.294), but DESIGNM does not adjust for a confounding variable, and that strong assumption of implicit matching is rarely justified in case-control studies, so this program was not made freely available. Software such as EGRET SIZ[10], which include sample sizes for conditional logistic regression, are alternatives to DESIGNM, which is based on Miettinen’s test of the Mantel-Haenszel odds ratio for matched case-control designs. The adjustment in DAYSMITH and DESIGN is for stratified matching [2, 11, 12], where matching is by confounders. This presumes that the eventual analysis will be unconditional [2] and will account for the stratification.

6. Interaction

The literature [1, 3] discusses interaction adjustment only for confounders with $K = 2$. It is easy, however, to modify the formulas for multi-level interaction. Every occurrence of R_E in the formulas (Appendix) is replaced by $R_E R_{I_j}$, where R_{I_j} is the interaction factor corresponding to the j^{th} level, $j = 2, \dots, K$. (For Σ' , put R_{I_j} inside the first sum.) We set $R_{I_1} = 1$.

For two confounder levels, R_{I2} , which is R_I in Smith and Day's notation [3], is the multiplicative factor by which the odds ratio for those exposed and in level 2 of the confounder is different from the odds ratio when there is confounder-exposure-disease interaction. For R_{Ij} , contrast is between level j and the reference level (level one).

This adjustment was made available for sensitivity analysis; specifically, to explore how much the sample size result could change if the confounder were in fact an effect modifier. Nevertheless, the adjusted formulas have been used to determine sample size in the presence of gene-environment interaction [13].

7. Discussion

The competitors to these programs are regression-based sample size programs, such as EGRET SIZ [10], which compute sample sizes required for unconditional or conditional logistic regression. The package nQuery [14] has an unconditional logistic regression option, but is not set up for case-control designs. These may be useful for continuous exposures, and make sense when the final analysis is intended to be such a regression, rather than a stratified analysis, such as a Mantel-Haenszel test, which our programs correspond to. We are unaware of any generally available competitor for stratified analysis.

In a series of papers on sample-size estimation to detect gene-environment interaction, which is a controversial role for sample-size formulas, comparisons have been made between regression based approaches and the stratified analysis approach [13, 15]. One solution is even to consider a case-only design [16]. EGRET SIZ does not provide for interaction adjustment.

When there is more than one confounder, we define one super-confounder, where each category corresponds to a sub-category. For example, if age, with 5 categories, and smoking, with 2 categories, are both confounders, then we define one super-confounder with $10 = 5 \times 2$ categories. The estimates of r_i , p_i , and R_{Ci} , $i = 2, \dots, 10$, then all have to take age and smoking into account jointly. As the number of confounders and the size of K increases, regression-based sample size programs become more advantageous, since information is not required for every sub-category.

The current programs yield results for 80% and 90% power, but versions are available for alternative powers, from 60% to 95%. A new version may print to the screen, if users want that option, and ask whether sample sizes for a specific power and Type I error are required.

The programs described are for two levels of disease (case vs. control) and of exposure. For several levels of exposure or disease, measures are available which correspond to odds ratios, risk ratios and risk differences [17], and it is not difficult to compute sample size formulas for these. If there is some demand, software to do those calculations may be created.

The Breslow-Day-Smith formulas which we extend utilize the classical method, based on testing. A more modern approach is that based on a confidence interval for the odds ratio [18], which may eventually become a program option. A Bayesian approach seems most suited for the sample size problem, although some issues need to be resolved [19]. Although not yet written, a Bayesian solution will soon be formulated for case-control designs.

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Appendix 1: The Formulas

For confounder adjustment in an unmatched design, to extend the formulas in [1], page 305, from one to F controls per case, we need only multiply the parameter Σ' in [1] by F .

We define $\Sigma' = F[PR_E \sum_{i=1}^K p_{1i}R_{Ci} + (1 - P) \sum_{i=1}^K p_{2i}R_{Ci}]$, and

$$V_A = (\sum 1/V_{iA})^{-1} \quad , \quad V_{iA} = \frac{1}{Pp_{1i}} + \frac{1}{(1 - P)p_{2i}} + \Sigma' \left[\frac{1}{Pp_{1i}R_{Ci}R_E} + \frac{1}{(1 - P)p_{2i}R_{Ci}} \right] \quad ,$$

$$V_N = (\sum 1/V_{iN})^{-1} \quad , \quad V_{iN} = T_i^3 / W_{1i}W_{2i}W_{3i}W_{4i}$$

(we divide out the factor n shown in [1]), where the proportion in confounder level i

$$\text{for a control is } W_{1i} = Pp_{1i} + (1 - P)p_{2i} \quad ,$$

$$\text{for a case is } W_{2i} = [Pp_{1i}R_E + (1 - P)p_{2i}]R_{Ci}/\Sigma' \quad ,$$

$$\text{and exposed is } W_{3i} = Pp_{1i}(1 + R_{Ci}R_E/\Sigma') \quad ,$$

$$\text{and unexposed is } W_{4i} = (1 - P)p_{2i}(1 + R_{Ci}/\Sigma') \quad ; \quad \text{and}$$

$$T_i = W_{1i} + W_{2i} = W_{3i} + W_{4i} \quad .$$

To compute the sample size, we invert formula (7.15) of [1], to obtain the one-sided ($S = 1$) solution

$$n = [(z_\alpha \sqrt{V_N} + z_{1-\beta} \sqrt{V_A}) / \log(R_E)]^2 \quad , \quad (1)$$

and the control sample size is the first integer greater than n . The case sample size is the first integer greater than n/F . The terms z_α and $z_{1-\beta}$ are the usual Standard Normal deviates. We replace z_α with $z_{\alpha/2}$ for the two-sided solution (when $S = 2$). Four common values of α and two values of β are supplied by the program.

The unadjusted formulas are shown on page 305 of [1], but it is easy to show that these are the same as the results obtained when we specify $K = 1$ in the preceding formulas. Note that $p_{11} = 1$ and $p_{21} = 1$ in that case. In the computer program, therefore, we use the same lines of computation for the adjusted and unadjusted results.

For confounder adjustment in a strata-matched design, The definition of Σ' is the same as above. The other items are re-defined as follows:

$$V_{iA} = \Sigma' \left\{ \left[\frac{1}{Pp_{1i}} + \frac{1}{(1-P)p_{2i}} \right] \frac{[Pp_{1i} + (1-P)p_{2i}]}{[Pp_{1i}R_E + (1-P)p_{2i}]R_{Ci}F} + \frac{1}{Pp_{1i}R_{Ci}R_E} + \frac{1}{(1-P)p_{2i}R_{Ci}} \right\},$$

$$V_{iN} = T_i^3 / W_{1i}W_{2i}W_{3i}W_{4i} = (1+F)^3 W_{2i} / FW_{3i}W_{4i}, \text{ with}$$

$$W_{1i} = FW_{2i},$$

$$W_{2i} = [Pp_{1i}R_E + (1-P)p_{2i}]R_{Ci} / \Sigma', \text{ as before,}$$

$$W_{3i} = Pp_{1i}R_{Ci}R_E / \Sigma' + FW_{2i}Pp_{1i} / [Pp_{1i} + (1-P)p_{2i}],$$

$$W_{4i} = (1-P)p_{2i}R_{Ci} / \Sigma' + FW_{2i}(1-P)p_{2i} / (Pp_{1i} + (1-P)p_{2i}),$$

$$\text{and } T_i = W_{1i} + W_{2i} = (1+F)W_{2i} = W_{3i} + W_{4i}.$$

Again, we use $V_A = (\sum 1/V_{iA})^{-1}$ and $V_N = (\sum 1/V_{iN})^{-1}$, which are the approximations due to Woolf [20].

To compute the sample size, we again use formula (1).